

## Neutrøns & Fond

Sydney | Australia 16-19 October 2018







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### Welcome to Neutrons and Food 5 and Sydney

On behalf of the local organising committee, it is my pleasure to welcome you to the 5th International Neutrons and Food conference and the beautiful city of Sydney, Australia.

This is the latest in a successful series of conferences on the application, science and technology of neutron scattering techniques as applied to food materials, first held in Sydney in 2010. The conference has been held every two years since with attendees showcasing how neutron scattering techniques have been used, often combined with other complementary measurement methods including X-ray and light scattering. I am privileged to have the opportunity to chair this event and that this conference has returned to Australia.

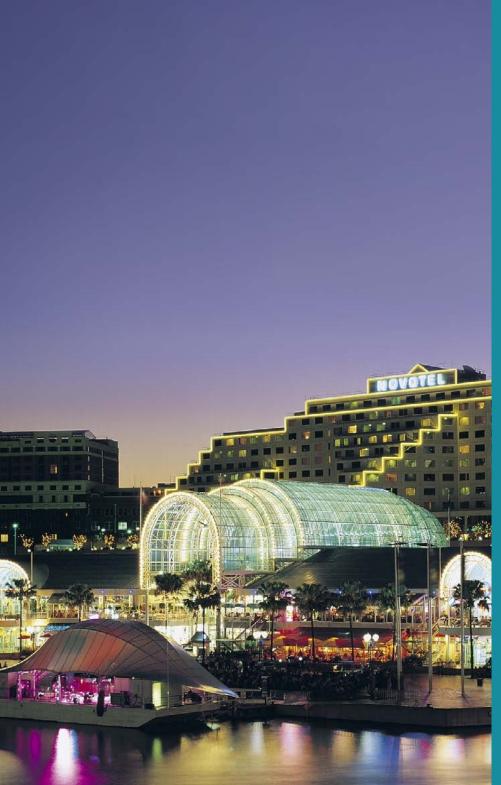
I would like to take this opportunity to welcome all of our delegates, especially those in the early stages of their career, some of whom will be showcasing their research at this meeting. I would also like to sincerely thank all of our sponsors; the conference would not have been possible without your generous support.

Neutrons and Food 5 offers all delegates a stimulating scientific programme that covers no fewer than ten themes addressing both traditional topics as well as emerging applications. With neutron scattering remaining at its core, we will also showcase nuclear-based techniques more broadly and how they are increasingly being used.

I wish to extend a special welcome to our overseas delegates. I appreciate that many of you have travelled a long distance to share your expertise with us and I hope that you enjoy your stay here in Sydney. I would finally like to welcome those delegates attending the Neutrons and Food conference for the first time. I hope that you find the meeting valuable and that this will represent the beginning of a regular event in your calendar.

I wish you all an extremely successful conference, both personally and professionally.

Elliot Gilbert Chair, Neutrons and Food 5







# Neutrons & Food 2016 | Sydney, Australia

## Overview

		DAY 1	DAY 2		DAY 3
Tuesday 16 October		Wednesday 17 October	Thursday 18 October		Friday 19 October
	<b>9.00</b> am		Food emulsions	<b>9.00</b> am	Plant materials
Outland	<b>10.00</b> am	Morning tea  Neutrons and Food Welcome		<b>10.00</b> am	
Optional tour ANSTO	<b>11.00</b> am	Glassy states  Processing, rheology and tribology	Morning tea  Lipids and fats	<b>11.00</b> am	Morning tea  Encapsulation and controlled
	<b>12.00</b> pm		Epido di la laco	<b>12.00</b> pm	release
	<b>1.00</b> pm	Lunch	Lunch	<b>1.00</b> pm	Lunch
Optional tour  Symbio Wildlife Park	<b>2.00</b> pm	Food packaging, preservation, safety and quality	Dairy	<b>2.00</b> pm	Industrial engagement Industry Q&A
	<b>3.00</b> pm	Afternoon tea	Afternoon tea	<b>3.00</b> pm	Neutrons and Food 6 Presentation
	<b>4.00</b> pm	Innovative methods for food (nuclear, synchrotron and light)	Innovative methods for food	<b>4.00</b> pm	
	<b>5.00</b> pm		(nuclear, synchrotron and light)	<b>5.00</b> pm	
	<b>6.00</b> pm			<b>6.00</b> pm	
	<b>7.00</b> pm		Dinner cruise	<b>7.00</b> pm	
Welcome Reception	<b>8.00</b> pm			<b>8.00</b> pm	
	<b>9.00</b> pm			<b>9.00</b> pm	

## Program

### Tuesday 16 October







OPTIONAL TOUR

Symbio Wildlife Park



7.30 pm Welcome Reception
Casa Risorante

2 h 🕓

### DAY 1

### Wednesday 17 October

<b>9.00</b> am	Registration	30 mins
<b>9.30</b> am	Morning tea	45 mins
<b>10.15</b> am	Neutrons and Food Welcome	25 mins 🕓
MORNING SES	SSION   Glassy states Gle	CHAIR b Yakubov   Bei Tian
<b>10.40</b> am	Importance and problems of water in food and food related materials  Jan Swenson - Chalmers University of Technology	40 mins 🕓
MID MORNING	s session   Processing, rheology and tribology	
<b>11.20</b> am	From wetting to wear: visualization of contact zone in beverage tribology <b>Stefan Baier</b> - <i>PepsiCo and University of Queensland</i>	20 mins 🕓
<b>11.40</b> am	Combining rheology and small-angle scattering of neutrons and X-rays for dynamic assessment of microfibrillated cellulose under shear Evgenii Velichko - TU Delft	20 mins 🕓
<b>12.00</b> pm	Neutron and X-Ray reflectivity from chocolate sandwiches Iva Manasi - University of Edinburgh	20 mins 🕓
<b>12.20</b> pm	Rheo-ND: Temperature and shear induced crystal transformation of a model triglyceride observed using neutron diffraction  Norman Booth - ANSTO	20 mins 🕓
<b>12.40</b> pm	Lunch - Sponsored by Riddet Institute	1 h 20 mins 🕓
MID AFTERNO	on session   Food packaging, preservation, Ramune Kuktaite   M safety and quality	CHAIR atthew van Leeuwen
<b>2.00</b> pm	Determining seafood provenance and safety through nuclear techniques  Debashish Mazumder - ANSTO	20 mins 🕔
<b>2.20</b> pm	The interaction of fluorinated compounds with a phospholipid bilayer <b>Shirin Nouhi</b> - <i>Uppsala University</i>	20 mins 🕓
<b>2.40</b> pm	Interaction of native and modified clupeine with Gram-negative model membranes  Marcia English - Saint Francis Xavier University	20 mins 🕓
<b>3.00</b> pm	Quality and stability evaluation of chicken meat treated with gamma irradiation and turmeric powder  Muhammad Arshad - Government College University Faisalabad, Pakistan	20 mins 🕓
<b>3.20</b> pm	Afternoon tea - Sponsored by ANBUG	30 mins
AFTERNOON S	ROBESSION   Innovative methods for food (nuclear, synchrotron and light)	CHAIR bbinson   Iva Manasi
<b>3.50</b> pm	Neutrons and Food - an introduction Elliot Gilbert - ANSTO	20 mins 🕓
<b>4.10</b> pm	Applications of Neutron Activation Analysis in food studies <b>Attila Stopic</b> - ANSTO	20 mins 🕔
<b>4.30</b> pm	Opportunities for QENS and neutron spectroscopy at ACNS <b>Nicolas de Souza</b> - ANSTO	20 mins 🕓
<b>4.50</b> pm	High-resolution macro ATR-FTIR chemical imaging capability at Australian Synchrotron IR Beamline and its applications in food science Jitraporn Vongsvivut - ANSTO	20 mins 🕓
<b>5.10</b> pm	X-ray fluorescence microscopy capabilities at the Australian Synchrotron and applications in food science  David Paterson - ANSTO	20 mins 🕔

### Thursday 18 October

MORNING SE	SSION   Food emulsions Tommy Nylande	CHAIR r   Lirong Cheng
<b>9.00</b> am	Lipid digestion – Key to the ability of milk-like emulsions to promote oral drug delivery  **Andrew Clulow - Monash University**	20 mins 🕓
<b>9.20</b> am	Oil-in-water emulsion system stabilized by emulsion droplets coated with whey protein microgels  Lirong Cheng	20 mins 🕓
<b>9.40</b> am	Interface structure of 70% fish oil-in-water emulsions stabilized with combinations of sodium caseinate and phosphatidylcholine: use of small angle neutron and X-ray scattering techniques  Betül Yesiltas - National Food Institute, Technical University of Denmark	20 mins 🕓
<b>10.00</b> am	Open Forum - Facilitator: Jamie Schulz	40 mins
<b>10.40</b> am	Morning tea - Sponsored by ACIS	40 mins
MID MORNIN	G SESSION   <b>Lipids and fats</b>	CHAIR Betül Yesiltas
<b>11.20</b> am	The memories of liquid triacylglycerols  Gianfranco Mazzanti - Dalhousie University	20 mins 🕓
<b>11.40</b> am	Structure formation in oleic acid - sodium oleate based oleogels <b>Steven Cornet</b> - Wageningen UR	20 mins 🕓
<b>12.00</b> pm	Static and dynamic multiscale characterisation of micronized fat crystal network formation and disruption by USAXS and rheo-SAXS  Tatiana Nikolaeva - Wageningen University	20 mins 🕓
MID MORNIN	G SESSION   Innovative methods for food (nuclear, synchrotron and light)	
<b>12.20</b> pm	Heavy food molecules from the National Deuteration Facility for structure function applications  Tamim Darwish - ANSTO	20 mins 🕓
<b>12.40</b> pm	Lunch - Sponsored by Office of NSW Chief Scientist	h 20 mins 🕓
MID AFTERNO	DON SESSION   Dairy Toshiji Kanay	CHAIR a   Shirin Nouhi
<b>2.00</b> pm	Structural characterization of milk coagulation from 0.1 to 20µm using Ultra-Small Angle Neutron Scattering  Alejandro Marangoni - University of Guelph	40 mins 🕓
<b>2.40</b> pm	Ultra-Small Angle Neutron Scattering investigation of milk coagulation: Data analysis and contrast matching methods  Nukhalu Callaghan-Patrachar - University of Guelph	20 mins 🕓
<b>3.00</b> pm	Dynamics of calcium caseinate in H <sub>2</sub> O or D <sub>2</sub> O studied by Quasi-Elastic Neutron Scattering (QENS)  Bei Tian	20 mins 🕓
<b>3.20</b> pm	Afternoon tea - Sponsored by <b>Anton-Paar</b>	30 mins
	Marcia English	CHAIR   Steven Cornet
<b>3.50</b> pm	Ultra-small angle neutron scattering studies on milk and cheese curd formation  Carl Adams - St. Francis Xavier University	20 mins 🕓
<b>4.10</b> pm	AStructure of Artificial Casein Micelles Composed of Deuterated β-Casein and Native κ-Casein: A Contrast Variation SANS and SAXS Study Jared Raynes - CSIRO	20 mins 🕓
AFTERNOON	session   Innovative methods for food (nuclear, synchrotron and light)	
<b>4.30</b> pm	Small Angle Neutron Scattering instrument BILBY: capabilities to study food science related problems  Anna Sokolova - ANSTO	20 mins 🕓
<b>4.50</b> pm	Neutron Imaging Application in Food Science on DINGO at OPAL <b>Ulf Garbe</b> - <i>ANSTO</i>	20 mins 🕓

### DAY 3

### Friday 19 October

MORNING SE	SSION   Plant materials Marianne Gaboriea	<b>CHAIR</b> u   Evgenii Velichko
<b>9.00</b> am	The potential of scattering techniques to investigate the structure and molecular interactions of Polysaccharides  Marta Martínez-Sanz - IATA, CSIC	40 mins 🕒
<b>9.40</b> am	Brush-like polysaccharides with motif-specific interactions Gleb Yakubov - University of Queensland	20 mins 🕓
<b>10.00</b> am	Supramolecular characterisation of starch in rice by NMR, SAXS and XRD Matthew Van Leeuwen - Western Sydney University	20 mins 🕓
<b>10.20</b> am	If we could design plant protein structures and tune properties in processed food Ramune Kuktaite - Swedish University of Agricultural Sciences	20 mins 🕓
10.40	Morning tea	40 mins
MID MORNIN	G SESSION   Encapsulation and controlled release Andrew Clulow   Nukhalu	<b>CHAIR</b> Callaghan-Patrachar
<b>11.20</b> pm	Neutron scattering study of lipid sponge-phase nanoparticles as enzyme carriers for food processing  Tommy Nylander - Lund University	20 mins 🕔
<b>11.40</b> pm	A combined SANS and USANS study to investigate the structure of solid lipid nanoparticles  Rohan Shah - Swinburne University of Technology	20 mins 🕔
<b>12.00</b> pm	Structural investigation on nanostructured lipid carriers for fish oil by small angle scattering  Martin Schmiele - Københavns Universitet	20 mins 🕓
<b>12.20</b> am	IUCr Travel Award Presentation	40 mins
<b>1.00</b> pm	Lunch - Sponsored by <b>ANSTO</b>	1 h 🕓
MID AFTERNO	oon session   <b>Industrial engagement</b>	CHAIR Jitendra Mata
<b>2.00</b> pm	Neutrons for industry - ACNS ILO, ANSTO - Access, services and case studies  Anna Paradowska - ANSTO	20 mins 🕓
<b>2.20</b> pm	Industry Q&A - Facilitator: Jitendra Mata	40 mins
<b>3.00</b> pm	Neutrons and Food 6 Presentation	20 mins 🕓



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## Invited speakers



Prof Alejandro Marangoni

University of Guelph

Prof Alejandro G. Marangoni is a professor and Tier I Canada Research Chair Food, Health and Aging at the University of Guelph, Canada.

Alejandro's work concentrates on the physical properties of lipidic materials in foods, cosmetics and biolubricants. He has published over 350 refereed research articles, 60 book chapters, 13 books, and over 40 patents. He is the recipient of many awards including the 2013 AOCS Stephen Chang award, the 2014 IFT Chang Award in Lipid Science, the 2014 Supelco/Nicholas Pelick Award, the 2015 ISF Kaufmann Medal and the 2017 Alton E. Bailey Award of the AOCS. Marangoni was honored as one of the 10 most influential Hispanic Canadians in 2012 and a Fellow of the American Oil Chemists' Society in 2015.

Alejandro is the first co-editor in Chief of Currer Opinion in Food Science, ElC of the Lipid Library and past Editor-in-Chief of Food Research International. Prof Marangoni has trained over 100 people in his laboratory; many occupy positions of importance in the academe and industry, including eleven professors at major



Prof Jan Swenson

Chalmers University of Technology

Jan was born in 1966 and earned his PhD in Physic at Chalmers University of Technology, Sweden, in 1996. He was thereafter a postdoctoral fellow at University College London, UK, the following two years, before he returned to Chalmers for an assistant professorship.

Jan was also employed by the Royal Swedish Academy of Sciences before he got a full professorship at NTNU, Norway, in 2005. Since 200 he has hold his current position as professor in physics at Chalmers. His current main research interest concerns the role of water in biological materials and how biological materials can be stabilized and cryopreserved by a cryoprotective substance such as trehalose

Jan is also working on the development of new simulation methods to model neutron scattering data. He has published about 160 reviewed scientific papers and has a H-index of 42



Dr Marta Martinez-Sanz

Spanish National Research Council (CSIC)

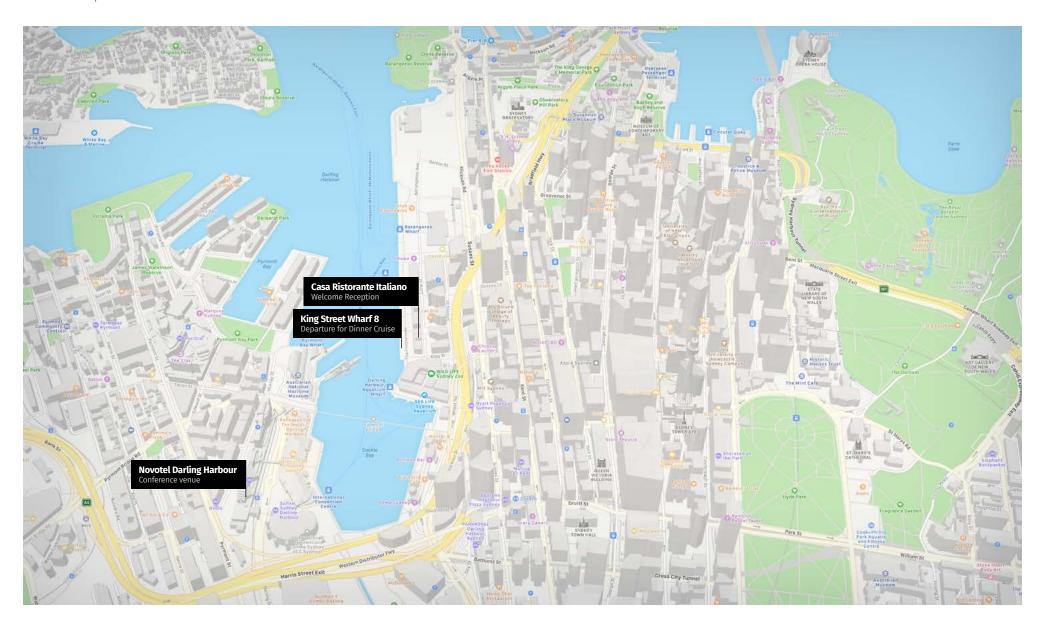
Marta received a BSc. (Hons) in Chemical Engineering and a Ph.D. in Food Science at the Polytechnic University of Valencia, Spain. She is currently a Juan de la Cierva fellow at the Food Quality and Preservation Department at the Institute of Agrochemistry and Food Technology (IATA-CSIC) (Spain).

Marta's research focuses on the extraction and characterisation of polysaccharides from renewable resources, with special focus on aquatic biomass, for the production of food additives and food packaging materials. Prior to this position, she worked for three years as a joint postdoctoral research fellow in the Australian Nuclear Science and Technology Organisation (ANSTO) and the University of Queensland (UQ). Marta's research focused on the investigation of the multi-scale structure of plant cell walls and model systems utilizing small angle scattering and diffraction techniques in combination with complementary microscopy, spectroscopy and rheology methods.

Marta carried out her PhD at the Institute of Agrochemistry and Food Technology (IATA,CSIC), specializing in the synthesis and characterisation of bio-based nanofillers extracted from renewable resources, such as plant-derived and bacterial cellulose nanocrystals and nanokeratin, as well as on the development of novel routes to incorporate them into polymeric matrices. Overall, her research is focused on the multi-scale structural investigation of bio-based materials and food-based systems, making use of a wide range of techniques and advanced characterization tools.

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### Map



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### Committees



### LOCAL ORGANISING COMMITTEE

Elliot Paul GILBERT	Kelly <b>CUBBIN</b>	Jitendra <b>MATA</b>
ANSTO	ANSTO	ANSTO
AUSTRALIA	AUSTRALIA	AUSTRALIA
Anna	Heidi	Ashley
PARADOWSKA	WELTON	HILL
ANSTO	ANSTO	ANSTO
AUSTRALIA	AUSTRALIA	AUSTRALIA



### **INTERNATIONAL ADVISORY COMMITTEE**

Vicky GARCIA-SAKAI	Amparo <b>LOPEZ-RUBIO</b>	Camille <b>LOUPIAC</b>
ISIS	IATA   CSIC	AgroSup Dijon
UNITED KINGDOM	SPAIN	FRANCE
John van	Anna	Alan
DUYNHOVEN	STRADNER	MACKIE
Unilever	Lund University	Leeds University
NETHERLANDS	SWEDEN	UNITED KINGDOM



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### Abstracts

### Importance and problems of water in food and food related materials

Most food contains relatively large amounts of water and this water is generally of central importance for the taste and other properties of the food. However, in the case of long-time storage this water can be problematic and cause a lot of detrimental aging effects. One way to overcome such aging effects is to reduce the temperature and thereby slow down the aging, but this gives often rise to ice formation, which generates other problems of damaged cellular structures in the food materials. To overcome this problem it is generally required to dry the food material before it is long-time stored at a low temperature, so called freeze-drying. Nevertheless, water in food materials, as well as

in living biological materials, is both essential and sometimes problematic. In this presentation these important and detrimental roles of water in carbohydrate rich food will be discussed. For example, we have elucidated the nature of ice crystals in frozen dough and bred [1], and how these ice crystals grow with incresing storage-time [2,3]. We also discuss attempts to reduce the detrimental ice formation. Furthermore, we have investigated how the dynamics of the water in carbohydrate rich food is affected by the structure and/or dynamics of the food material. It was found that the water dynamics becomes slower in food materials with slow dynamics, which implies that the dynamics of the water and the pure food material are interrelated [4]. It was also evident that the water dynamics speeds up dramatically with incresing water content [4]. Finally, the anomalous dynamical properties of deeply supercooled water will be discussed, and how this affects the freeze-storage of biological materials.

- [1] G. Chen, H. Jansson. K. F. Lustrup and J. Swenson. Formation and distribution of ice upon freezing of different formulations of wheat bread. J. Cereal Sci. 55, 279 (2012).
- [2] G. Chen et al. Long-term frozen storage on the dynamics of water and ice in wheat bread, I. Cereal Sci. 57, 120 (2013).
- [3] J. Eckardt et al. Long term frozen storage of wheat bread and dough Effect of time, temperature and fibre on sensory quality, microstructure and state of water. J. Cereal Sci. 57, 125 (2013).
- [4] H. Jansson, W. S. Howells and J. Swenson. Dynamics of fresh and freeze-dried strawberry and red onion by quasielastic neutron scattering. J. Phys. Chem. B 110, 13786 (2006).

### **AUTHOR**

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### From wetting to wear: visualization of contact zone in beverage tribology

Matching the mouthfeel of sugar is still one of the largest unsolved challenges of product develop- ers, as sugar not only provides sweetness, but also mouthfeel. Mouthfeel is one of the most critical product attributes driving consumer choice and acceptability of reduced sugar beverages. In order to link mouthfeel attributes to analytical measures, one must first of all appreciate that drinking is a highly dynamic process that involves mixing of the ingested beverage with saliva and the resulting bolus being sheared over several length scales between the tongue and the upper palate. In order to better interpret saliva driven changes to beverage boli, it is important to recognize the conversion of the beverage from a simple Newtonian fluid to a multiphase complex fluid. Currently, our ability to properly interpret tribology data from multiphase complex fluids is rather limited, as we lack additional information on structure (atomic, molecular, macromolecular) in the contact zone. Therefore developing techniques that will allow us to measure structural changes governed by saliva beverage interactions will lead to better insights to enable the rational design of reduced sugar beverages.

The authors are employed by PepsiCo, Inc. The views expressed in this abstract are those of the authors and do not necessarily reflect the position or policy of PepsiCo, Inc.

### **AUTHORS**

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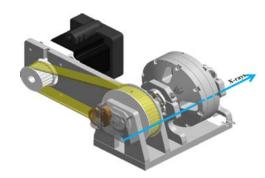
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# Combining rheology and small-angle scattering of neutrons and X-rays for dynamic assessment of microfibrillated cellulose under shear

Cellulose fibre is a natural food component present in fruits and vegetables. Presence of the cellulose fibre in the diet aids digestion and helps in preventing a variety of diseases. Cellulose fibres also provide texture to food products. The texturing capacity of cellulose fibres can be enhanced by defibrillation under high shear. Dispersions of microfibrillated cellulose form a 3D soft space-filling network, which can, for example, mimic the texture of fat-based products. Critical for the consumer perception of microfibrillated cellulose networks are spreadability and mouthfeel. The microstructural features that underly these rheological parameters are poorly understood. Small-angle scattering of neutrons (SANS) and X-rays (SAXS) provides structural information in a wide range of length scales. Combination of rheological data with SANS/SAXS measurements of the samples under shear conditions sheds light on the mesoscale structural dynamics that underlie the peculiar flow of microfibrillated cellulose under industrial processing conditions and during consumer use and consumption.

Recent studies [1] have shown nonlocal flow behaviour of microfibrillar cellulose suspensions. Based on Rheo-MRI data two different hypotheses to explain such nonlocality were suggested: flocculation of cellulose microfibrils and formation of a liquid-crystal like phase in these suspensions. To validate these hypotheses we constructed a special sample cell, which allows for SANS/SAVS on the materials under shear with spatial resolution (100  $\mu$ m) across the velocity gradient direction of the cell. No alignment of microfibrills was observed at shear rates from 1 to 300 s-1. The scattering invariant calculation indicates a homogeneous density distribution of cellulose microfibrils across the gap at all applied shear-rates. This implies that no shear-induced aggregation and concomitant migration of cellulose over the gap occurs. The shear-SAXS experiments are in line with the flocculation hypothesis with the flock sizes much larger than 200nm (maximal accessed in the performed experiment).

[1] D. W. Kort et al., Soft Matter, DOI: 10.1039/c5sm02869h (2016).



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### Neutron and X-Ray reflectivity from chocolate sandwiches

Chocolate is a dense suspension of solids, mainly sucrose, in a continuous triglyceride fat phase of cocoa butter, containing Lecithin, which is mostly phospholipid, and in some cases the polymeric surfactant PolyGlyceryl PolyRicinoleate (PGPR). These surfactants reduce the yield stress and viscosity of molten chocolate, which is important in chocolate manufacture and influences the mouth feel of the chocolate. How these surfactants cause these modifications to the rheology of molten chocolate remains an open question.

We have developed a methodology that allows the preparation of well-defined crystalline planar sucrose films as substrates to study the structures formed by these surfactants at sucrose/triglyceride interfaces. X-ray and neutron reflectivity has been used in combination with QCM-D to characterize the adsorption of these surfactants individually and in combination at the sucrose/triglyceride interface to provide a structural basis for the observed rheology.

[1] Birgit Schantz, Harald Rohm. (2005) Influence of lecithin–PGPR blends on the rheologi- cal properties of chocolate. Lebensm.-Wiss. u.-Technol. 2005, 38: 41-45. http://www.sciencedirect.com/science/article/pii/S0023

Keyword: Rheology, interfacial structure, reflectivity, triglyceride

### **AUTHOR**

Iva **Manasi** 

### **CO-AUTHOR**

Simon Titmuss

1 University of Edinburgh



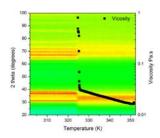
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### Rheo-ND: Temperature and shear induced crystal transformation of a model triglyceride observed using neutron diffraction

Rheo-SANS (rheology and small angle neutron diffraction) is now a wellestablished technique to probe the shape and size of particles under different shear and temperature regimes 1. Recent work on WOMBAT 2, the high intensity neutron diffraction instrument at the Australian Centre for Neutron Scattering, has successfully combined rheology and neutron diffraction for the first time. This was used to follow the crystalline phase transformations in a model (deuterated) triglyceride.

The initial impetus for this work was part of a forensic investigation [3] linking the crystallisation of triglycerides under high shear rates that are encountered in motor vehicle accidents. However now the technique has been demonstrated it may be of interest to investigate fat crystallization due to shear in food manufacture. Possible applications might include investigation of detrimental crystallisation during conching or tempering of chocolate or extrusion of foods high in fats (string cheese?).



The high neutron flux and detector efficiency available at WOMBAT is instrumental in allowing diffraction patterns to be collected from the small sample volumes available in the Couette cell geometry. The Couette cell used has a sample gap of 1mm and was mounted tangentially to the neutron beam. The shear rate applied to the sample can be varied over a range of 10-2500 s-1 and the temper- ature can be controlled using an external water jacket from between -10 and 80 0C. We invite other researchers who are interested in this capacity to contact us about possible experiments.

- [1] Tabor, R.F., Zaveer, M.I., Dagastine, R.R., Grillo, I and Garvey, C.J., Phase Behavior, Small-Angle Neutron Scattering and Rheology of Ternary Nonionic Surfactant-Oil-Water Systems: A Com- parison of Oils, Langmuir 29(11), 3575-3582 (2013).
- [2] Studer, A.L., M.E. Hagen, and T.I. Noakes, Wombat: The high-intensity powder diffractometer at the OPAL reactor, Physica B: Condensed Matter, 2006, 385-386, Part 2(0): p. 1013-1015.
- [3] Stuart B, Thomas P, Maynard-Casely H, Booth N, Leung A, A neutron diffraction investigation of shear forces on a model lipid for forensic application, ANZFSS 23rd International Symposium on the Forensic Sciences, September 2016, Auckland.

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### Determining seafood provenance and safety through nuclear techniques

Demand for farmed or wild-caught seafood is growing globally. Regulatory bodies, market chain actors and consumers have an interest in seafood provenance and food safety assessment tools to protect human health, guarantee sustainability, to meet certification requirements, reduce or prevent food fraud, and to minimise biosecurity risks associated with global movement of raw seafood products. The limitations of current seafood provenance methods can be overcome using complementary

or standalone nuclear techniques. Food quality is a complex term and has often been associated with food safety by consumers; traceability, a key requirement of the global seafood trade, is linked to both food safety and quality. This study investigated traceability and food quality using nuclear techniques. The traceability component focused on determining the geographical origin and production method of Asian seabass (Lates calcarifer) and black tiger prawns (Penaeus monodon) from a range of Asia-Pacific locations using Stable Isotope Analysis (SIA) and X-ray fluorescence (XRF) through Itrax. The food quality component focused on using Neutron Activation Analysis (NAA) to detect the concentration of heavy metals and other elements (39 in total) in the two species.

The data were analysed using three different statistical methods; univariate and multivariate anal- ysis, Linear Discriminant Analysis (LDA), and randomForest. Both LDA and randomForest had consistent results: XRF effectively distinguished the production method and geographic locations of P. monodon (up to 100% accuracy), and SIA was more effective for L. calcarifer (up to 94% accuracy). In most cases for both L. calcarifer and P. monodon, NAA revealed that the concentrations of heavy metals and micronutrients were below the threshold levels of concern for human consumption. There were no significant differences between farmed and wild-caught samples or between geographic locations. However, in some cases, elements such as arsenic and selenium were signifi- cantly higher than the threshold, but their chemical forms are yet to be determined.

SIA and XRF are effective complementary methods for determining the provenance of seafood. NAA should be combined with other methods such as protein analysis and fatty acid profiling to effectively determine the quality of seafood.

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Neutrons & Food 2018 | Sydney,

### The interaction of fluorinated compounds with a phospholipid bilayer

Some industrial surfactants that are commonly used in preparation of non-stick coatings for kitchen dishes and in food packaging products are fluorinated compounds as they have good water and stain repellent properties. These materials however now attract increasing global concern due to their persistence, bioaccumulation and possible adverse effects on the environment and living organisms.

Concerns regarding these molecules being carcinogenic and possibly harmful for children immunes system have been raised. The use of some of these compounds, in particular long-chain compounds, has been banned in many countries and some other compounds are under discussion as regards continued use. In the past few years, new alternative molecules with shorter chain length or with partly fluorinated chains have been introduced as replacements that are thought to be safer products.

There is however little known about the interaction of these compounds with biological systems or well-founded reasons for specific choices. In this project, we have investigated the interaction of fully and partly fluorinated compounds with model systems relevant to biological interfaces using neutron reflectometry. Neutrons provide a unique and powerful tool to investigate the amount and location of fluorocarbons when mixed with hydrogenous molecules.

The study covers a range of compounds, from the ones that are already banned to new replacement alternatives It has allowed a comparison of the effect of hydrophobic chain length and of different hydrophilic functional groups on their interactions with phospholipid bilayers. Fully fluorinated molecules have been shown to penetrate into bilayers and displace the lipids whereas partly fluorinated compounds show negligible interactions with the lipid. Off-specular scattering data from bilayers which have been contacted with fully fluorinated compounds indicate that there are rough and patchy structures after exposure. Extensive rinsing with water removes some of the fluorocarbon surfactant but a less dense lipid bilayer is left behind.

The interaction of these compounds has been found to vary with the head group as: sulfonamide > sulfonate > carboxylic acids and to increase with the chain length [1] which strongly correlates with their solubility in water.

[1] S. Nouhi et al. (2018), Journal of Colloid and Interface Science, 511, 474-481.

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### Interaction of native and modified clupeine with Gram-negative model membranes

Clupeine, a cationic antimicrobial peptide found in the sperm cells of fish, is of interest as a food additive because of its antimicrobial activity against several foodborne pathogens. However, it has previously been shown that non-specific binding of clupeine to anionic molecules reduces its antimicrobial activity. It has also been shown that the overall positive charge of the native peptide can be reduced by blocking 10% of its arginine residues with 1,2-cyclohexanedione (CHD) to form CHD-treated clupeine. CHD-treated clupeine retains antimicrobial activity but it is not known if the modes of interaction against Gram-negative bacteria remain the same as the native peptide. The focus of the present study was to investigate the effect of charge reduction on peptide membrane interactions by comparing the effect of native and CHD-treated clupeine on Escherichia coli (E. coli) model biomembranes.

It was hypothesized that the reduction in charge would result in different interactions with model monolayers composed of E. coli PE, phosphatidylethanolamine, 79 mole%:PG, phosphatidylglycerol, 17 mole%: and CL, cardiolipin, 4 mole%), and model bilayer membranes composed of DPPC,: PE:PG:CL (79:17:4 mole%).

Peptide interaction with the model biomembranes were studied using Neutron Reflectometry (NR) and X-ray reflectometry (XRR) and symmetric bilayers were deposited on silicon blocks applying the Langmuir-Blodgett and Langmuir Schaefer techniques. Some lipid mixing was observed in the inner tail region (~69 ± 0.24% DPPC (1.2-dipalmitoy) (d62)-snglycero-3-phosphocholine) and ~24 ± 0.02% PE:PG:CL); and in the outer tail region (~24 ± 0.02% DPPC and ~56 ± 0.01% PE:PG:CL). Native and CHD-treated clupeine were not able to cross the model PE:PG:CL:DPPC bilayer biomembrane, however, CHD-treated clupeine showed increased interactions with the lipid head group. Inspite of the different interactions observed in the model systems, a more comprehensive understanding of the safety and toxicology of both peptides is required before they can be used for food applications.

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# Neutrons & Food 2018 | Sydney, Austra

# Quality and stability evaluation of chicken meat treated with gamma irradiation and turmeric powder

The safety of chicken meat increased by treatment of gamma radiation. The free radicals produced due to radiation in food products. To overcome this issue, some source of antioxidants used along with radiation. Turmeric powder has the ability to work as antioxidant as well as antimicrobial. The study was carried out to evaluate the impact of gamma irradiation and turmeric powder (TP) on microbial quality, physicochemical quality, stability and antioxidant status of chicken meat. There were 2 doses (1 kGy and 2 kGy) of gamma irradiation was used alone and with combination of 3% turmeric powder along with control (0kGy). Aerobic and vacuum packaging were used for the storage of chicken meat with interval of 0,7 and 14 days at 4°C.

The microbiological results showed that the level of contamination was in the decreasing order as the dose of gamma irradiation increases both for total bacteria and coliforms, whereas no contamination was found in group treated with 2 kGy+TP both for aerobic and vacuum packaging. The peroxide value (POV), thiobarbituric acid reactive substances (TBARS) and total volatile basic nitrogen (TVBN) differed significantly on chicken meat with different groups. Higher POV and TBARS were found in chicken meat treated with 2 kGy under aerobic packaging after 14 days of storage and TVBN was higher in control at day 14 of storage under aerobic packaging whereas minimum POV, TBARS and TVBN were found in 0kGy+TP at day 0 under vacuum packaging.

The results showed that different sensory attributes were evaluated by the panel of judges and higher score for all the sensory attributes like appearance, taste, texture, flavor and overall acceptability was found in 2 kGy. It is concluded that the chicken meat treated with 2 kGy+TP was considered better for microbial and physicochemical quality, antioxidant activity as well as sensorial properties of chicken meat.

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### Neutrons and Food - an Introduction

The application of neutron scattering methods to understand the structure and dynamics in soft- condensed matter has a long history but its utilisation for food-based materials still remains the domain of a small, albeit increasing, community. This is despite the fact that neutrons offer significant advantages over other avenues of characterisation where an understanding at the nanoscale is essential to explain the functional properties and behaviour in food. Neutrons are isotopically sensitive and are also highly penetrating enabling transmission through complex sample environments; the latter provides the potential to study industrially-relevant processes in real time. The presence of higher flux facilities, with access to extended spatial and energy ranges, greater computational power and enhanced modelling methods, offer further opportunities. This presentation will provide a brief introduction to neutron scattering and how such methods can yield useful information on food materials.

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# Neutrons & Food 2018 | Sydney, Austr

### Applications of Neutron Activation Analysis in food studies

Neutron Activation Analysis (NAA) is a sensitive technique for quantifying elemental concentrations in solid samples. It is a mature and well-understood method that has been in use for several decades across a range of disciplines. Today it is one of many techniques available to analysts for characterising the elemental makeup of solid samples, albeit with somewhat limited availability as a powerful neutron source, such as a nuclear research reactor, is a requirement. Partly but not entirely as a result of this, it is often overlooked in favour of more readily accessible methods, yet still holds some key advantages over other methods commonly used. This talk aims to cover the basic concepts of NAA and outline the advantages and disadvantages in the context of elemental analysis of food, while outlining the experience of the NAA capability at the Australian Nuclear Science & Technology Organisation (ANSTO).

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## Opportunities for QENS and neutron spectroscopy at ACNS

The Australian Centre for Neutron Scattering presently operates two neutron inelastic spectrometers well suited to quasi-elastic neutron scattering (QENS) and low-energy neutron spectroscopy studies. In combination, the EMU and PELICAN cold-neutron spectrometers cover an energy transfer range

spanning 0.001 to 14 meV. As far as QENS, the accessible time domain is thus from pico- to nano- seconds, with a momentum transfers ranging from 0.1 to beyond 2  $\mathring{A}^{-1}$ .

The main features of the two instruments will be presented, including their available sample environments. EMU is a backscattering spectrometer, and PELICAN is a polarised time-of-flight spectrometer. Examples highlighting the capabilities of each spectrometer and their complementarity will be presented, with a focus on application areas such as soft-matter and bio-material relaxations, and aqueous/hydration dynamics.

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### **High-resolution macro ATR-FTIR** chemical imaging capability at **Australian Synchrotron IR Beamline** and its applications in food science

This work presents advances in high-resolution chemical imaging capability at Australian Synchrotron Infrared (IR) beamline, achieved through the use of an in-house developed synchrotron macro ATR- FTIR microspectroscopic device [1]. The device was developed by modifying the cantilever arm of a standard macro-ATR unit to accept germanium (Ge) ATR elements with different contact facet sizes (i.e. 1 mm, 250 µm and 100 μm in diameter). Coupling synchrotron-IR beam to the Ge ATR element (n=4) used in this device, has the effect of not only reducing the beam focus size (improving the lateral resolution) by a factor of 4, but also reducing the mapping step size by 4 times relative to the stage step motion. As a result, the macro ATR-FTIR measurement at Australian Synchrotron IR Beamline can be performed at minimum beam size of 1.9 µm using a 20x objective, and at minimum mapping step size of 250 nm, allowing high-resolution chemical imaging analysis.

While the large Ge facet size works well with soft materials that do not require high pressure to achieve a good contact, the small tips can provide higher pressure and allow measurements inside smaller regions with limited access suitable for hard and rough surfaces [1-2]. This macro ATR-FTIR device can also be coupled to a temperature control unit, allowing temperature-dependent study, as well as measurements that require a fixed temperature such as analysis of dairy products at 4°C similar to the usual storage condition in a household fridge.

The development of the macro ATR-FTIR device has so far led to successful analysis of samples from a diverse range of research disciplinary. Key applications in food science to be presented include a range of dairy products (e.g. cheese and yoghurt), plants and vegetables.

- [1] V.K. Truong, M. Stefanovic, S. Maclaughlin, M.J. Tobin, J. Vongsvivut, M. Al Kobaisi, R.J. Crawford, E.P. Ivanova, "The evolution of silica nanoparticle-polyester coatings on surfaces exposed to sun-light." I. Vis. Exp. 116, e54309, 1-11 (2016).
- [2] J. Vongsvivut, V.K. Truong, M.A. Kobaisi, S. Maclaughlin, M.J. Tobin, R.J. Crawford, E.P. Ivanova, "Synchrotron macro ATR-FTIR microspectroscopic analysis of silica nanoparticle-embedded polyester coated steel surfaces subjected to prolonged UV and humidity exposure," PLoS One 12, e0188345 (2017).

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### X-ray fluorescence microscopy capabilities at the Australian Synchrotron and applications in food science

X-ray fluorescence microscopy (XFM) can be used for elemental and chemical microanalysis across length scales ranging from millimeter to nanometer. XFM is ideally suited to quantitatively map trace elements within whole and sectioned plants, seeds, animals and soil samples. The high elemental sensitivity of X-ray fluorescence microprobes coupled with deep penetration of hard X-rays enables measurement of an incredibly diverse range of samples in situ and under environmental conditions with a minimum of preparation.

The ability to rapidly acquire 2D images enables higher-dimensional studies such as fluorescence tomography, X-ray absorption near edge structure (XANES) imaging, and XANES tomography in realistic times. Full spectral XANES imaging takes advantage of fast XFM and results in X-ray absorption near edge structure spectra from X-ray fluorescence at each pixel in the image. This enables spatially resolved chemical speciation and the efficiency and speed ensures the lowest possible dose alongside high

Examples of XFM elemental mapping addressing bio-fortification of cereal grains and chemical speciation studies to understand manganese and arsenic toxicity in plants and selenium bio-availability in aquatic environments will be presented.

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# Neutrons & Food 2018 | Sydney, Australi

### Lipid digestion – Key to the ability of milk-like emulsions to promote oral drug delivery

Milk is a staple of the human diet and is an essential nutrient source for all mammalian infants. The fat content of milk comprises 98% triglycerides emulsified with milk fat globular membrane material and native proteins. Digestion of these triglycerides in the intestines yields monoglycerides and fatty acids that are absorbed through the intestinal wall. Lipophilic molecules such as drugs and nutrients can be dissolved in the milk fat globules but fat digestion can result in further dissolution or precipitation of incorporated drugs and nutrients. Digestion of milk fats also leads to the formation of intricate liquid crystalline structures that evolve throughout digestion,[1,2] which may also affect the release of incorporated drugs and nutrients. The process of lipid digestion is therefore key to the fate of lipophilic molecules dissolved in the fat globules of the milk emulsion and directly controls milk's ability to act as an excipient for lipophilic drugs and nutrients.

In this presentation, small angle X-ray and neutron scattering techniques (SAXS and SANS, respec- tively) will be used to probe the structural transformations occurring during the digestion of milk fats and the subsequent effects on incorporated drugs. Synchrotron SAXS reveals that the liquid crystalline structural transformations occurring during the digestion of milk fats are robust with respect to heat treatment, freezing and powdering/reconstitution of milk.[3] It will also be established that milk-substitutes such as infant formula and vegetable juices do not necessarily replicate milk-like structures during digestion. The distribution of digestion products within fat globules at different extents of digestion will be examined with SANS combined with selective deuteration of milk digestion products. Finally, the incorporation of the amphiphilic antimalarial drug OZ439[4] into milk and infant formula will be outlined and the influence of lipid digestion on the solubility and bioavailability of OZ439 will be discussed.

- [1] Salentinig et al. Formation of Highly Organized Nanostructures during the Digestion of Milk. ACS Nano 2013, 7 (12), 10904-10911.
- [2] Salentinig et al. Self-Assembly Structure Formation during the Digestion of Human Breast Milk. Angew. Chem. Int. Ed. 2015, 54 (5), 1600-1603.
- [3] Clulow/Salim et al. A closer look at the behaviour of milk lipids during digestion. Chem. Phys. Lipids 2018, 211, 107-116.
- [4] Clulow et al. The Curious Case of the OZ439 Mesylate Salt: An Amphiphilic Antimalarial Drug with Diverse Solution and Solid State Structures. Mol. Pharm. 2018, in press, DOI:10.1021/acs.molpharmaceut.8b00173

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### Oil-in-water emulsion system stabilized by emulsion droplets coated with whey protein microgels

Structurally designed emulsions are a developing group that is likely to find increasing utilization within the food industry because of their potential advantages over conventional emulsions. A novel dropletstabilized (DS) emulsion system emulsified with casein micelles has been previously reported1. However, the mechanism of DS emulsion formation. physicochemical properties, and their stability are not yet been fully explored. In the present study, heat-induced whey protein microgel (WPM) particles were used as an alternative emulsifying agent. The structure of WPM particles on the formation and physicochemical properties of the primary (PE) and the DS emulsions was investigated. WPM particles were prepared by heating 4 wt% whey protein isolate solution in the presence (PB) or absence (NPB) of 10 mM phosphate buffer at pH 5.9, 85°C for 45 min, followed by washing, centrifugation, and micro-fluidization. The PE coated with WPM were homogenized using 3 passes at the pressure of 250/50 bar. DS emulsions were prepared by mixing (at 30000 rev/min for 2 min) 10 wt% oil with 10, 30 or 60 wt% PE. The structure of WPM particles and emulsions were analyzed by dynamic light scattering, confocal light scattering microscopy (CLSM), transmission electronic microscopy (TEM), and the combination of small and ultra-small angle neutron scattering (SANS and USANS).

The results showed that the WPM particles produced in the absence of phosphate buffer (WPM-NPB) were smooth spherical particles, giving a surface fractal dimension of 2.0 and a hydrodynamic diameter of 270 nm. However, WPM particles made in the presence of phosphate buffer (WPM-PB) were rough spherical particles with a surface fractal dimension of 2.3 and a hydrodynamic diameter of 290 nm. Particle fragments present in the WPM-PB dispersion, resulted in their competitive adsorption onto the surface of the DS emulsions; reducing the adsorption of PE droplets. For the PE coated with WPM-NPB particles (PE-NPB), a flocculation due to protein bridging and protein intramolecular interaction, lead to a network with a fractal dimension of 2.7. For the DS emulsions stabilized by PE-NPB. the interfacial layer thickness of DS emulsion droplets increased with the increase in the concentration of PE as observed by CLSM, whereas the size of DS emulsion droplets decreased. A fractal network consisting of adsorbed PE-NPB on the interfacial layer of DS emulsion was observed by TEM and measured by USANS. These results suggest that both the structure of the interfacial layer and the size of the DS emulsion is dependent on the concentration of the PE used.

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# Interface structure of 70% fish oil-in-water emulsions stabilized with combinations of sodium caseinate and phosphatidylcholine:

use of small angle neutron and X-ray scattering techniques

Marine long chain (LC) omega-3 polyunsaturated fatty acids (PUFAs) have been reported to have numerous beneficial effects on health such as decreasing cardiovascular diseases, improving immune system and mental health. Oil-in-water emulsions have been used as delivery systems to en- rich foods with LC omega-3 PUFAs, such as eicosapentaeonic (C20:5n-3, EPA) and docosahexaeonic (C22:6n-3, DHA) acids, in order to increase the intake of these bioactive compounds. However, LC omega-3 PUFAs are highly prone to oxidation, which results in formation of lipid oxidation prod- ucts causing undesired sensory properties as well as loss in nutritional profile. As high fat omega-3

delivery emulsions are applicable to many food systems and lipid oxidation in emulsions have been claimed to be initiated at the oilwater interface, food researchers and industry are highly interested in understanding the inner dynamics of these systems. This study focused on characterizing interfacial structure of high fat (70%) oil-in-water emulsions emulsified with sodium caseinate (CAS) and phosphatidylcholine (PC). Structure of emulsifiers which are adsorbed on water-oil interface could be best studied by Small Angle Neutron Scattering (SANS) together with Small Angle X-Ray Scattering (SAXS). Interfacial structure was aimed to be characterized by identifying thickness of the interfacial layer and distances between emulsifiers adsorbed at the oil-water interface as well as obtaining information on adsorption behavior of the combined use of emulsifiers in high fat delivery emulsions. In order to study these characteristics, simpler versions of this system was also studied; e.g. different concentrations of CAS in D<sub>2</sub>O, different concentrations of PC in D.O. 70% oil-in-water emulsions produced only with CAS or only with PC at different concentrations. Contrast matching between oil and water phase as well as contrast variation of water phase were applied. For this reason, scattering length densities and relative concentrations of employed compounds were calculated and hydrogen content was considered to be minimized. From these considerations, it was proposed to have a model system of deuterated water and various mixtures of deuterated and regular hexadecane or only deuterated hexadecane providing maximum contrast between liquids and emulsifiers. Fish oil was also included in some of the measurements in order to compare the results from model systems with the original system.

SAVS results have shown that CAS forms aggregates and PC forms multilayers both in water and high fat oil-in-water emulsion. It was found that CAS concentration affected the periodic repeat distance of the PC bilayers. When the mass concentration of CAS increased, the distance between PC bilayers become smaller, this was attributed to the decrease in water amount trapped between PC layers. When the concentration of CAS was fixed, the distance between PC bilayers was the same independently from PC concentration. Moreover, SANS results indicated that the aggregate size was bigger when CAS was in the emulsion interface together with PC compared to the CAS in D<sub>2</sub>O which might be an indication of the interaction between CAS and PC at the oil-water interface in high fat oil-in-water emulsions.

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### The memories of liquid triacylglycerols

Crystallization of triacylglycerols (TAG), and other lipids, happens from the liquid into a broad range of polymorphic crystalline forms. Despite the sustained research effort over many years, it is still not possible to predict the crystalline form from first principles. In the case of mixtures, the prediction becomes disproportionately hard, even for just binary mixtures. They exhibit a very large number of seemingly anomalous crystallization behaviours. Among them, the 'memory effect' is particularly relevant. Perhaps the liquid structure holds the key to these phenomena. A brief review of the few conceptual models proposed for the organization of TAG molecules in the liquid shows that, until a few years ago, researchers had not been able to offer a solid proposal for the liquid structures. Yet, the hypothetical structures suggested in the literature are often taken for granted.

We present here results from experiments aimed at proposing a structure of the liquid state and its influence on crystallization phenomena that are otherwise hard to explain. Essentially, TAG molecules form clusters due to the difference in attractive forces between two regions of the molecules: the aliphatic chains, and the polar glycerol core. The clustering has been predicted by molecular dynamics and coarse-grained simulations.

Pure liquid TAG samples were examined by x-ray scattering at temperatures up to 210°C. Wide an- gle scattering (WAXS) data are consistent with the liquid phase of aliphatic molecules. Small angle scattering (SAXS) data are similar to those produced by alcohols and fatty acids, whose molecules associate via polar groups. The liquid TAG seem to form "Loose Multimers" of 5 to 9 molecules. The average number of molecules per cluster decreases with temperature and increases with molecular weight.

The 'memory effect' is observed when TAG re-crystallize from a liquid, obtained by melting crystalline TAG, and form the same structure that they had as a solid. Differential scanning calorimetry (DSC) experiments were done with tempered pure TAG and TAG mixtures. The materials were melted and held at many [time + temperature] combinations before recrystallizing. Combinations that produced a recrystallization equal to a crystallization from a random liquid defined a time-temperature boundary. Above the boundary, the memory is erased. For pure TAG, the times and temperatures required were shorter and lower than for the mixtures. Some liquid structure of pure triacylglycerols remains after melting, which is disrupted by thermal fluctuations. In blends, it is additionally necessary to homogenize the concentration of domains of different sizes, via molecular diffusion, a much slower process.

Ultra-small angle neutron scattering (USANS) experiments were performed using tripalmitin. Data obtained from hydrogenated and deuterated samples were collected at 95°C. Data from the deuter- ated material were then obtained upon cooling from the melt, and upon melting from the crystalline state. The difference in the scattering patterns indicates the presence of different structural organization in both cases, consistent with the explanation of the 'memory effect' due to differences in the clustering of molecules in the liquid state.

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### NTS $\stackrel{\frown}{=}$

## Structure formation in oleic acid - sodium oleate based oleogels

The structuring of liquid oil into what are known as oleogels has received considerable attention in recent years. Oleogels have the potential to replace solid fat in food products with healthier unsaturated oils. Various oleo-gelators are known, ranging from large polymers to low molecular weight oleo-gelators (LMWOG). A known combination of LMWOGs is that of oleic acid and sodium oleate. Their ability to gel sunflower oil has already been demonstrated, and gel strength can be tuned by varying the ratio between oleic acid and sodium oleate. The addition of a small amount of water results in stronger gels and suggests hydrogen bonding plays a role in the gelation. [1]

In this presentation we will dive deeper into the relation between the gel strength of oleogels prepared with oleic acid and sodium oleate, and their micro- and nano-structure. Oleogels were pre- pared with different ratios of the two LMWOGs and using different triglyceride oils. Upon varying the ratio between oleic acid and sodium oleate, changes in both micro- and nano-structure were ob- served through light microscopy and Small Angle Neutron Scattering (SANS) and Ultra-SANS (US- ANS). The observed structure ranges from inverse micelles to lamellar crystals, which coexist in some cases. Additional information on the type of crystals was collected using Small Angle X-ray Scattering. In addition to the triglyceride oils, n-hexadecane was used as a solvent analogue. This enabled the use of selective deuteration and provided additional information on the nano-structure. A clear relation between micro- and nano-structure, and the strength of the oleogel was observed.

[1] RSC Adv.,2015, 5, 47466

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### Static and dynamic multiscale characterisation of micronized fat crystal network formation and disruption by USAXS and rheo-SAXS

Micronization of fat crystals presents an appealing alternative for the conventional melt-cool routes for manufacturing fat based food products. By dispersing micronized fat crystals (MFC) in oil, fat crystallisation and network formation can be decoupled, which can bring significant process singular process the industrial application scope of these dispersions we carried out a multiscale investigation on MFC network formation and disruption by means of USAXS, rheo-SAXS, confocal Raman imaging and rheo-MRI.

Rheo-SAXS and rheo-MRI showed that upon dispersion in oil, MFC network formation was concomitant with recrystallisation. Oil type, temperature and shear rate collectively determined MFC recrystallisation rate, which inversely correlated with the strength of the resulting weak-link network where crystal aggregates are embedded in a continuous net of crystalline nanoplatelets. US-AXS revealed that the rough surface of MFC nanoplatelets hampers stacking into one-dimensional aggregates ('TAGwoods'), which explains the high mass fractal dimension of the networks formed in MFC dispersions as compared to those formed by melt-cooling.

Applying shear to matured MFC networks leads to a gradual and irreversible loss of yield stress, as shown by rheo-MRI. Shear did however not affect network fractal dimensions (USAXS) and also did not disrupt micronscale MFC aggregates (confocal Raman imaging). Rheo-SAXS revealed that loss of network strength can be attributed to release of nanoplatelets from the weak-link network, which subsequently align in the shear field and undergo rapid recrystallisation. Our insights in the factors that govern MFC network formation and disruption bear relevance for simplified manufacturing of fat-based food products by effectively turning their design into a colloidal aggregation game.

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# Neutrons & Food 2018 | Sydney, Australi

# Heavy food molecules from the National Deuteration Facility for structurefunction applications

There have been limited global initiatives in the field of molecular deuteration where the majority of these programs focus on biological deuteration of proteins, while more complex deuterated small molecules like phospholipids, sugars, triglycerides, fatty acids haven't been widely available to the wider science community. This has limited the experiments that can be performed, and formed a bottle-neck for advancing the applications of neutron scattering, mass spectroscopy and NMR.

In this paper we will discuss the recent advancements and the impact of deuteration on the research outcomes achieved by using deuterated molecules produced by the National Deuteration Facility of the Australian Nuclear Science and Technology Organisation. Recent high-impact case studies in the fields of food science and biotechnology will be presented which reveal the exciting and diverse characterisation studies which are now available for the neutron, NMR, IR and mass spectroscopy science communities.

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### Structural characterization of milk coagulation from 0.1 to 20µm using Ultra-Small Angle Neutron Scattering

Milk is a main component in many food products in a variety of cultures around the world. Research and development in milk over the past five decades has enabled increased shelf-life, better flavor and accurate control over fat size and concentration. Milk is a water-based colloidal system, comprised of soluble proteins, casein micelles (CMs), ions and fat globules (FGs). The addition of a coagulant, the enzyme chymosin, to this system brings about aggregation of the CMs via hydrolysis of the κ-casein molecules which make up the surface of the CMs, effectively trapping the FGs together with some liquid whey, resulting in a complex soft matter system called cheese. The structure and organization of the CMs and FGs in milk have been addressed by many invasive experimental methods probing scales from 0.1nm to 1µm, yielding conclusions via two-dimensional images. Furthermore, the structure of cheese probed by these invasive methods (CLSM, cryo-SEM, etc.) could introduce artifacts to the unperturbed structure. In view of this, it is desirable to utilize a technique that can be non-perturbing and can access the micron-scale structure of cheese.

This technique is ultra-small angle neutron scattering (USANS), which enables us to probe length scales from 0.1µm to 20µm, effectively providing information on the size, surface characteristics, threedimensional organization, scattering length density and concentration of the main components in milk and cheese. Our USANS studies were carried out on the BT-5 thermal neutron beam port at the NIST Center of Neutron Research, Gaithersburg, Maryland. Scattering results from homogenized milk revealed smooth-surfaced FGs with mean diameters of 540nm, polydispersity of 40% and concentration of 3.28%. Utilizing the contrast matching technique with homogenized milk samples and overlapping the signal with scattering data from skim milk, CMs were found with mean diameters of 120nm, polydispersity of 16% and concentration of 1.5%. When homogenized milk was coagulated with chymosin for one hour, CMs were found to be organized in one-dimensional aggregates having an average thickness of about two CMs. Correlations of coagulation time with aggregate thickness and surface roughness displayed aggregate sizes doubling (for two hours), then slightly decreasing (for three hours), while the surface became smoother as coagulation time increased. During skim milk coagulation, smooth aggregates of two CM thickness formed quickly (within 1 hour), continuing to double in thickness while becoming rougher (within 2 hours) - possibly attributable to the absence of FGs.

Our results suggest that the USANS technique is a promising method to probe micron-scale structure of commercial milk in natura, as well as milk coagulated via enzymatic activity, since our results pertaining to diameter and concentration are in agreement with those in the literature. Pros and cons of this technique with regard to applications in milk and cheese studies will be highlighted and data analysis methods discussed. A comparison to the USAXS technique will also be offered. Characterization of mean sizes, polydispersity's and independence in the organization of CMs and FGs, can contribute to optimization in the manufacture of healthy milk-based food products.

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## Ultra-Small Angle Neutron Scattering investigation of milk coagulation:

Data analysis and contrast matching methods

Milk, a water-based colloidal system, is comprised of fat globules (FGs), casein micelles (CMs), soluble proteins and ions. The mechanism of cheese formation from milk, through the induced aggregation of CMs by the addition of chymosin, is a complex soft matter process. This process has been widely characterized using invasive experimental methods (confocal microscopy, electron mi- croscopy, etc.) that have probed milk and cheese at length scales from 0.1nm to 1µm, providing two-dimensional images of the organization and structure of the main components in milk, viz. CMs and FGs. Given that these techniques could impose artifacts on the system, in the present study we utilized ultra-small angle neutron scattering (USANS) to investigate the coagulation of homogenized whole (HW) and skim milk in natura. The USANS instrument probes length scales from 0.1µm to 20µm, providing information on size of the clusters, surface characteristics, three-dimensional organization, scattering-length density (SLD), polydispersity and volume fractions of the main components in the milk and cheese samples. An advantage of using neutrons is the ability to utilize contrast matching, a technique that varies the scattering-length of the hydrogen-deuterium component in the system to effectively blend certain objects (CMs or FGs) into the background, so as to observe a clean signal from the object of interest. Raw USANS data can be analyzed using a variety of models to describe the micron-structure of the scattering objects. We have implemented two analytical models, Guinier-Porod (GP) and Bimodal Shulz-Spheres (BSS). These models provide us with the most persuasive interpretation of the process of milk coagulation. Through examination of the slopes using the GP model, we found correlations of coagulation time with the CM aggregate thickness and surface roughness, as well as the shape or packing symmetry of the aggregate. Analysis using the BSS model enabled us to identify the polydispersity, volume fraction, SLD and av- erage radii of FGs and CMs in HW milk. With this model it is possible to decompose the combined scattering effects from two objects (assumed to be spheres) within a liquid medium, while taking

the variability in radii of the polydisperse objects into account. Contrast matching confirmed the observed three-phase system by running a series of HW milk samples diluted with different ratios of D2O to H2O using the predicted SLDs and given volume fractions. Blending of the FGs in HW milk (3.25 wt% milk fat) with the background signal proved successful when compared to the skim milk (< 0.5 wt% milk fat) data, as it showed little to no difference (Figure1). Our findings demonstrate the value of USANS in combination with GP and BSS modeling techniques as analytical tools for the characterization of the micron-structure of commercial milk in natura, as well as coagulated milk obtained via enzymatic action.

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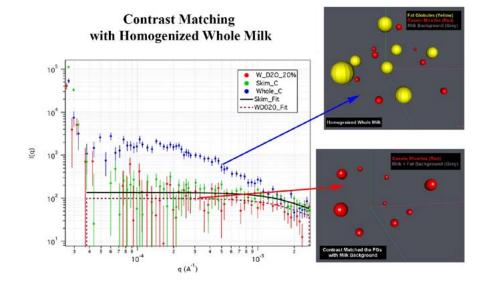
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Producing meat analogues using calcium caseinate is an intermediate step to reduce carbon emission and improve animal welfare. Depending on the drying methods of the powder, 30% w/w spray-dried calcium caseinate (SCaCas) forms pronounced fibrous structure under shear and heat of  $50^{\circ}$ C, while roller-dried calcium caseinate (RCaCas) does not [1]. Solvent plays a role in the fibrous structure as well. When SCaCas is prepared with D<sub>2</sub>O, the formation of fibrous structure is not possible while RCaCas shows anisotropy in the final structure [2]. We hypothesize that drying methods and solvent environments affect the hydration behaviour of the protein powders.

To systematically study the dynamics of calcium caseinate, we performed QENS experiments on both SCaCas hydrated with  $H_2O$  (SH) or  $D_2O$  (SD) and RCaCas with  $H_2O$  (RH) or  $D_2O$  (RD). Samples with a hydration level of 0.4g/g dry powder were measured with an energy resolution of -17.5  $\mu$ eV and  $O_2$ -range of  $O_2$ -17. Å-1.

Mean Square Displacements (MSDs) were extracted by fitting the elastic window scans with Gaussian approximation and is shown in figure 1. The 'dynamical transition' temperature of the SH is the highest (250K), and its slope above 250K the steepest. Results suggest, for RCaCas, the activation energy for protein dynamics is insensitive to solvent. While for SCaCas, the protein dynamics is the same as RD when hydrated with D<sub>2</sub>O, but more energy is needed to induce dynamical transition when hydrated with H<sub>2</sub>O. Plus, the water dynamics of SH is more active comparing to the rest.

The full energy spectra are fitted with the sum of a Delta function, a Lorentzian function and a linear background. The Full Width Half Maximum (FWHM) of the Lorentzian is fitted with the SingwiSjölander model to describe the motions of water molecules. The translational diffusion coefficient (D<sub>v</sub>) and residence time ( $\tau_0$ ), plotted in figure 2, were compared between samples at 293K (room temperature) and 320K (50°C). The D<sub>v</sub> of both samples increased with increasing temperature, with RH displaying a slightly higher increase (from 0.15 to 0.30 Å2/ps, compared to 0.17 to 0.27 Å2/ps for SH). As for  $\tau_0$ , both samples decreased with increasing temperature. In contrast to bulk water, whose  $\tau_0$  is typically 1ps, the water molecules are quite confined to the protein surface, even at elevated temperature. The difference between SH and RH may be attributed to the differences in initial powder morphology and conformational changes in structure at a higher temperature.

- [1] J.M. Manski, A.J. van der Goot, and R.M. Boom, "Formation of fibrous materials from dense calcium caseinate dispersions", Biomacromolecules, vol. 8, no. 4, pp. 1271-1279, 2007
- [2] B. Tian, ZJ Wang, AJ. van der Goot, and W.G. Bouwman, "Air bubbles in fibrous caseinate gels investigated by neutron refraction and X-ray tomography", Food Hvdrocolloids. in press.

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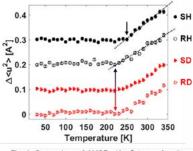
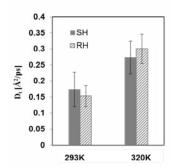


Fig. 1. Comparison of  $\Delta$ MSDs ( $\Delta$ < $^{QS}$ ) as a function of temperature.  $\Delta$ MSD is the difference between the hydrated powder and dry powder. The SD, RH and SH are shifted 0.1, 0.2 and 0.3  $A^2$  respectively for better visualization. The dash lines and arrows are guides for the eyes, indicating the slope and onset temperature of the dynamical transition.



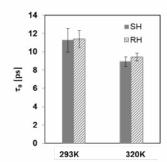


Fig. 2. Comparisons of changes in translational diffusion coefficient  $D_t$  (left) and residence time  $\tau_0$  (right) at 293K and 320K between the two H<sub>2</sub>O hydrated samples.

# Neutrons & Food 2018 | Sydney, Australi

### Ultra-small angle neutron scattering studies on milk and cheese curd formation

Milk and milk products are essential to global nutrition and the world-wide food industry. We have carried out ultra-small angle neutron scattering (USANS) studies on samples of "as-is" commercial food-grade milk and samples with added rennet, as the first step in cheese formation. Casein protein micelles are present in both skim and whole milk with median diameters of 120 nm with 16% polydispersity. In homogenized milk (3.25% milk fat) the median diameters of fat globules are 0.54 µm with a polydispersity closer to 40%. Despite various structural features implied by microscopy (rough kappa casein molecules, porosity, a membrane layer for fat globules, co-aggregation of micelles and fats) both of these components can be modelled by smooth, independent, uniform spheres. Contrast matching gives scattering length densities (SLD) of 0.20  $\times$  10<sup>-6</sup> Å<sup>-2</sup> for fats and 0.89  $\times$  10<sup>-6</sup> Å<sup>-2</sup> for casein micelles. A typical protein SLD is twice this value so the micelles are likely guite hydrated. Rennet dramatically increases the scattering at the smallest angles, consistent with the formation of large structures. As the curd making process continues, we observe further aggregation.

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## Structure of artificial casein micelles composed of deuterated β-casein and native κ-casein:

A contrast variation SANS and SAXS study

Milk is one of the most important commodities is the world and yet the colloidal protein structure, the casein micelle, that gives rise to many of the unique properties of milk is still not well defined due to its complex and dynamic structure. To gain a detailed picture of the structure of the casein micelle, we produced a world-first model artificial casein micelle comprising recombinant deuterated, phosphorylated,  $\beta$ -casein and native  $\kappa$ -casein, a micelle composition not dissimilar to "human" casein micelles. Using contrast variation SANS and SAXS, a detailed structure of the casein micelle will be presented.

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# Neutrons & Food 2018 | Sydney, Austral

# Small Angle Neutron Scattering instrument BILBY: capabilities to study food science related problems

Australian Nuclear Science and Technology Organization (ANSTO) for more than ten years successfully operates first Small Angle Neutron Scattering instrument QUOKKA [1] and in January of year 2016 commenced user operation of the second SANS instrument, BILBY [2]. The inspiration for BILBY is the D33 instrument located at the Instituteue-Langevin [3]. Similar to D33, BILBY exploits neutron time-of-flight to extend the measurable Q-range, over and above what is possible on a conventional reactor-based monochromatic SANS instrument.

BILBY has been designed to operate in two different modes: monochromatic and time-of-flight (ToF) mode, where four choppers are used to create neutron pulses. The latter gives a wide range of choice in relation to the wavelength resolution (from -3% – 30%, depending on the chopper setup and on the sample-detector distance). Two arrays of position sensitive detectors in combination with utilizing of wide wavelength range (from -2Å to -20Å) provide capability to collect scattering data of wide angular range without changing experimental set-up (the most common settings used by now allow simultaneous data collection in the range between 110-3Å-1 and 0.6Å-1, with the highest accessible angle -1.8Å-1).

The question is how the advanced design features can be applied to the real world of complex systems, like food. In short, having large Q-range available in one go, open up a possibility to study complex system, like a food-related gels, micelles, hierarchical features at the large scale of sizes without changing the instrument set-up. An option to tune the resolution allows one to resolve structural features very close in dimensions so the majority of the existing SANS machine cannot distinguish them. Or, in contrary, for the samples lacking defined structural features, relaxing resolution allows to increase neutron flux decreasing time of the data collection.

Also, additionally to having an instrument flexible in set-up by itself, there is a range of sample environment devices available for users. It provides, for example, a possibility to change the temperature of a sample, or apply the shear in-situ, or mix a complex system' component in the beam collecting data simultaneously, which is priceless for study food related samples.

BILBY is the new instrument which is servicing users for under three years, but already has a portfolio which demonstrates the benefits of features briefly describes above.

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### Neutron imaging application in food science on DINGO at OPAL

The new neutron radiography / tomography / imaging instrument DINGO [1] is operational since October 2014 to support research at ANSTO. It is designed for a broad national and international scientific user community and for routine quality control for defense, industrial. cultural heritage and archaeology applications. With experience from materials science and archeology we would like to connect to the field of food science application. DINGO provides a useful tool to give a different insight into objects because of different contrast compared to X-rays and high sensitivity to light elements. Since being operational we gathered experience in various scientific fields, with industrial applications and commercial customers demanding beam time on DINGO. The measured flux (using gold foil) for an L/D of approximately 500 at HB-2 is 5.3 x 107 [n/cm<sup>2</sup>s] allows us to run neutron tomography experiments in a reasonable time scale from 3hours to 3days per tomography, depending on sample composition and resolution. A special feature of DINGO is the in-pile collimator position in front of the main shutter at HB-2. The collimator offers two pinholes with a possible L/D of 500 and 1000. A secondary collimator separates the two beams by blocking one beam and positions another aperture for the other beam. The neutron beam size can be adjusted to the sample size from 25 x 25 mm<sup>2</sup> to 200 x 200 mm<sup>2</sup> with a resulting pixel size from 13µm to ~100µm. The whole instrument operates in two different positions, one for high resolution and one for high speed. We would like to present example experiments with potential new applications in food science.

[1] Garbe, U; Randall, T; Hughes, C; Davidson, G; Pangelis, S and Kennedy, SJ (2015), A New Neutron Radiography / Tomography / Imaging Station DINGO at OPAL, Physics Procedia 69, 27-32.

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# Scattering to characterizer sample preper strongly affer application is specificity at

# The potential of scattering techniques to investigate the structure and molecular interactions of polysaccharides

Scattering techniques represent an excellent tool for the structural characterization of natural polysac- charides since they involve minimal sample preparation and do not require drying processes which can strongly affect the native structure of hydrated systems. However, the application in this field is still a largely unexploited area due to their specificity and the complexity of data manipulation and interpretation.

In this talk, the application of small angle X-ray and neutron scattering (SAXS and SANS) as well as wide angle X-ray scattering (WAXS)/X-ray diffraction (XRD), in combination with complementary methods such as differential scanning calorimetry (DSC), spectroscopy and morphological characterisation, to investigate the structure and molecular interactions of polysaccharides in hydrated systems with different practical applications, such as the following: (i) plant cell wall (PCW) materials, including model hydrogels based on pure cellulose and composites with PCW polysaccharides (arabinoxylan, xyloglucan, mixed linkage glucans and pectins), mature cotton fibres and food-extracted PCWs; (ii) starch-microalgae aqueous blends for the development of functional foods and biodegrad- able packaging materials; (iii) gelatin/i-carrageenan encapsulation structures for the controlled release of bioactive compounds.

These examples highlight the potential of small angle scattering techniques to provide valuable insights on the structure and molecular interactions of polysaccharides in a wide variety of hydrated systems.

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## Brush-like polysaccharides with motif-specific interactions

Polysaccharide hydrocolloids are a class of biological polymers with unique physicochemical properties, versatile functionality and a broad spectrum of applications in foods, where they are used as thickening, gelling and structuring agents. Unlike proteins and DNA, typical food polysaccharides comprise a relatively small repertoire of constituent monomers, making their properties difficult to modulate without a need for chemical modification.

Here we report the discovery and rheological and structural characterisation of a special class of polysaccharides found in seed mucilage of plants from the Plantago genus. These polysaccharides have a xylan backbone densely decorated with an array of side-chains. Almost 95% of xylose resides of the backbone are decorated with side-chains, which classifies these polymers as bottle-brushes. Typically, the side-chains of these polymers are small oligosaccharides, 1 to 4 residues long, and comprise arabinose and xylose, and glucuronic acid residues. The variety of possible permutations of residues within side-chains confers these polysaccharides a number of unique aspects, including motif-specific interactions.

In particular, we have isolated two gel-forming arabinoxylan (AX) fractions by solvent specific ex- traction of Plantago ovata seed mucilage: AX-A(alkali) and AX-W(water) [1-3]. Both fractions are found to be neutral bottle-brush polysaccharides exclusively comprised of xylose and arabinose with similar molecular weight and linkage composition. Despite compositional similarity, their rheolog- ical and structural properties are markedly different. Using a combination of USANS/SANS with

rheological characterisation techniques, we have established the hierarchical nature of hydrogel assembly, with pore sizes in the range between 30 and 1000 nm. Using enzymatic cleavage of terminal arabinoses of side chains we induced significant changes in rheological and structural properties of these complex arabinoxylans, which enabled us to corroborate that (i) hydrogen bonding between side chains of neighbouring molecules is a key driver of gel formation; (ii) the strength of hydrogen bonding is motif specific, which explains differences in physical properties between two fractions. It is envisaged that discovery of motif-dependent interactions will opens new opportunities for rational design of polysaccharides with targeted and highly tuned physical properties, providing a far-reaching potential for delivering novel tailor-made hydrocolloids for use in foods, pharmaceutical industry, and smart materials.

- [1] Yu,L,Yakubov,G.E.,Zeng,W,Xing,X.,Stenson,J.,Bulone,V.,&Stokes,J.R.(2017).Multi-layer mucilage of Plantago ovata seeds: Rheological differences arise from variations in arabinoxylan side chains. Carbohydrate Polymers, 165, 132-141.
- [2] Yu, L., Yakubov, G. E., Martínez-Sanz, M., Gilbert, E.P., & Stokes, J. R. (2018). Rheological and structural properties of complex arabinoxylans from Plantago ovata seed mucilage under non-gelled conditions. Carbohydrate Polymers, 193, 179-188.
- [3] Yu, L., Yakubov, G. E., Gilbert, E.P., & Stokes, J. R. (2018). Multi-scale assembly of hydrogels formed by highly branched arabinoxylans from Plantago ovata seed mucilage studied by US- ANS/SANS and rheology. Carbohydrate Polymers, in press.

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# Neutrons & Food 2018 | Sydney, Australia

## Supramolecular characterisation of starch in rice by NMR, SAXS and XRD

Rice has fed more people over a longer period of time than any other grain [1] and currently provides 85 % of the energy intake to over half the world's population [2]. This makes it a suitable vehicle to help manage weight and obesity-related illnesses, especially type 2 diabetes and colorectal cancer.

Starch is the major component of rice (-90 %) and its supramolecular structure is known to affect rice digestibility. However, starch is one of the most complex materials found in nature with six known hierarchical levels of structure [3]. In rice, starch granule heterogeneity and its influence on digestibility depends upon genetic makeup, environmental influences (between geographical locations and seasons) and processing (such as milling and cooking).

Multiple levels of starch structure in cooked and raw rice were characterised to better understand how starch structure affects digestibility. Long range crystallinity was characterised by powder X-ray diffraction (XRD). Smaller scale crystalline structure in the form of helix content was assessed by 13C solid-state nuclear magnetic resonance (NMR) spectroscopy. The semi-crystalline lamellar structure of starch in rice was characterised by small angle X-ray scattering (SAXS) [4]. Complementary to SAXS, small angle neutron scattering (SANS) is a powerful technique in the analysis of soft condensed matter [5]. Sample versatility and solvent contrast makes SANS especially attractive in the analysis of granular starches under conditions relevant to food such as cooking [6]. Molecular mobility was also assessed by 1H solid-state NMR, determining the T2 relaxation time of starch in cooked rice.

This work aims to characterise the structural features of starch in a range of rice varieties, and to relate this information to rice digestibility. It is expected that the supramolecular structure of starch in rice will have a significant influence on the digestion of rice.

- [1] J. Zuxun, Q. Weifen, Y. Seo, J. Darby, R. Bowman, in Crop Post-Harvest: Science and Technology, Blackwell Science Ltd, 2007, pp. 1-25.
- [2] G. P. Hettel, J. L. Maclean, D. C. Dawe, Rice almanac: source book for the most important economic activity on earth, CABI Pub., New York, 2002.
- [3] M. Gaborieau, P. Castignolles, Caractérisation de l'amidon et de ses matériaux composites. Les An- nales des falsifications de l'expertise chimique et toxicologique (Société des Experts Chimistes de France) 2009, 9710, 23-32.
- [4] J. Blazek, E. P. Gilbert, Carbohydrate Polymers 2011, 85, 281-293.
- [5] A. Lopez-Rubio, E.P. Gilbert, Neutron scattering a natural tool for food science and technology research. Trends in Food Science & Technology 2009. 20, 576-86.
- [6] J. Doutch, M, Bason, F. Franceschini, K. James, D. Clowes, E.P. Gilbert, Structural changes during starch pasting using simultaneous Rapid Visco Analysis and small-angle neutron scattering. Carbohydrate Polymers 2012, 88, 1061-71.

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## If we could design plant protein structures and tune properties in processed food

Plant proteins from pea, wheat and quinoa during latest years have become attractive alternatives to replace imported soya in various food applications in Europe. Proteins in a form of protein-rich fractions (from pea and wheat), and the whole flour (from quinoa) can be highly attractive ingredients in making pasta, protein-rich snacks and various types of breakfast food. Structure, state of aggregation and morphology of protein components is a key factor determining of their functional properties in these foods and is greatly impacted by the processing method used. Therefore, a better control and understanding is needed on how protein structure and function are related in various protein containing systems of the processed food.

Here, we show few examples on structural-function relationship of the processed pea, wheat and quinoa proteins into pasta-like sheets, textured protein snacks and breakfast food. We studied the

protein structure and protein interactions (including components, as dietary fiber) using synchrotron WAXS/SAXS, X-ray scattering tomography. SEM and HPLC techniques. Variation in processing conditions was high and low processing temperature, composition of a blend and a processing method used. The results indicated that pea protein fraction in a blend with pea fiber showed polymerization behaviour that was greatly depended on the protein to fiber ratio (more protein greater polymerization). An increase of dietary fiber in the blend with pea protein resulted into high amounts of unordered structures observed by FTIR and in an increase in the distances of the scattering elements observed by SAXS. The mechanical properties, as strength and extensibility, of the pea protein and fiber blends were the highest for greatest amounts of protein having blends, while the E-modulus was similar for all the studied blends. For wheat protein fraction, gliadin, the structure and polymerization of gliadin textured snack were influenced by the composition of the blend as was observed by X-ray scattering tomography. Large microstructural differences, mainly due to high processing temperature treatment, were observed in the quinoa extrudates studied by SEM and X-ray scattering tomography.

Here we conclude, that understanding on how to control structure on macro- and nano- levels is a key element when tuning functional properties for the diverse protein systems in food. Once the protein structure is better understood, it would allow better steering of processing and control of final end-use characteristics of the protein rich food products.

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# Neutron scattering study of lipid sponge-phase nanoparticles as enzyme carriers for food processing

Immobilization of enzymes into different support materials has been widely used as means to control their activity and stability. This has mainly been done in context of bioanalytical, preparative or biomedical purposes. In the present study we investigate two key types of enzymes used in food processing, namely Aspartic protease (34 KDa) and Beta-galactosidase (460 KDa), which are used in processing of dairy products.

Mostly these proteins are delivered into the process as solutions with a considerable amount of preservatives and still with limited shelf-life and limited control of the enzyme activity. Here we have used lipid liquid crystalline phases as enzyme carriers, based on their established capability for drug delivery, protein encapsulation or crystallization. They can form a wide range of self-assembled structures with aqueous cavities of nano-scale dimensions. Reverse cubic or hexagonal lipid aqueous phase can be used to entrap smaller biomolecules yet it is still challenging to encapsulate bioactive macromolecules, such as proteins. Here, we will present a novel lipid system able to form highly swollen sponge phases (L3), with aqueous pores up to 13 nm of diameter.

We will show that this structure is preserved even if in excess aqueous solution, where they form sponge-like nanoparticles (L3 NPs) in which the two enzymes are included. The structure and composition of the particles was revealed by combined measurements using small angle neutron scattering (SANS), light scattering, cryo-TEM, size exclusion chromatography and Raman spectroscopy.

The SANS results reveal differences in the L3 NPs with and without enzyme. To reveal the nature of the interaction between the enzymes and the lipid matrix, we further studied the adsorption of both proteins on the lipid layers formed by the L3 NPs. These data reveal partial penetration of the enzymes in the lipid bilayers. The results of this study will be discussed in terms of the ability of these nanoparticles to encapsulate and release of the proteins in the lipid matrix.

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# A combined SANS and USANS study to investigate the structure of solid lipid nanoparticles

Solid lipid nanoparticles (SLNs) have been extensively investigated as drug carrier systems since their inception in the 1990s. These are emulsions when prepared at high temperature and crystallize predominantly into SLNs upon cooling. The details of their ultrastructure are poorly defined. Previously, our group reported a novel microwave-assisted microemulsion-based technique to prepare SLNs with radii of approximately 150 nm.

To understand the detailed internal structure of these SLNs, contrast variation ultra-small angle neutron scattering (USANS) and small angle neutron scattering (SANS) experiments were conducted on suspensions of hydrogenated stearic acid SLNs in D20. Together, SANS and USANS gave a combined Q range of 0.000047 to 0.6 Å  $^{\rm A}$ –1 (corresponding to a size range of 1 nm  $^{\rm +15}$  µm). The extended Q range used in this study allowed an extensive study of the hierarchical structure of SLNs.

The combined data are consistent with the SLNs having an oblate structure at the microscale level, intermediate between rods and lamellae, with roughened surfaces. At the nanoscale level, the results were consistent with the SLNs having an ellipsoidal shape intermediate between spheres and rods, with a crossover from mass fractals to surface fractals. The elucidation of this structure is particularly important given that the structure influences the stability and drug release properties of the nanoparticles. These results will assist in the development of systems with desired shape and properties.

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### Structural investigation on nanostructured lipid carriers for fish oil by small angle scattering

Fortification of foods with essential functional lipids such as  $\omega$ -3 polyunsaturated fatty acids (PU-FAs), β-carotene and vitamin A are of key importance for health and well-being. ω-3 PUFAs are essential for the human diet to support a good brain and cardiovascular health. Furthermore, they lower the risk of other diseases such as type 2 diabetes and inflammatory disorders. Vitamin A and its precursor β-carotene are important for growth and development, good vision and immune functions. Integrating such chemically unstable compounds into food products and delivering them into the human body requires encapsulating them into a carrier system. This improves their water-dispersibility and protects them from lipid oxidation.

Recent studies [1-3] show that lipid oxidation of ω-3 PUFAs reduced significantly when incorporated into a solid tristearin matrix, a so called nanostructured lipid carrier, when sufficient amounts of tristearin is added to the samples and lecithin with a high phase transition temperature and bile salts or Quillaja saponins are used as emulsifiers. From this observation it is anticipated that during the cooling process the emulsifier molecules at the lipid-water interface promote a co-crystallization of the lecithin chains with the tristearin via heterogeneous interfacial nucleation. The crystallized tristearin limits the mass transport of reactive agents which reduces lipid oxidation of the encapsulated fish oil.

However, the structural composition of such nanostructured lipid carrier particles is still unclear. Does the crystallized tristearin form a shell around a single fish oil core or is the fish oil embedded as smaller islands in the tristearin particle matrix?

Typically, these lipid particles are about 150 nm in size as measured by dynamic light scattering. Based on the sample composition (fish oil: tristearin ratios between 60:40 and 20:80) and assuming a spherical core-shell structure, we can expect thicknesses between 10 and 30 nm for a tristearin shell [3]. Thus, small-angle X-ray and neutron scattering (SAXS, SANS) and wide-angle X-ray scattering are the methods of choice to study the morphology of the particles and especially their internal structure on atomic to colloidal length scales.

In SANS and SAXS experiments we will study samples with different fish oil: tristearin ratios. Using mixtures of water and deuterated water for the dispersion medium as well as mixtures of tristearin and its fully deuterated analogue, three different neutron scattering contrasts are realized for each sample. The different contrasts allow to highlight the spatial distribution of both fish oil and tris-tearin alone and as a combination in the lipid particles.

After an introduction to the subject, the presentation will show first analysis results of the SAXS and SANS measurements with regard to the structural composition of the nanostructured lipid carriers containing fish oil.

[1] Salminen et al., Food Chem. 190 928-937 (2016)

[2] Salminen et al., Food Chem. 141(3) 2934-2943 (2013)

[3] Salminen et al., I. Colloid Interface Sci. 490 207-216 (2017)

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### Neutrons for industry - ACNS ILO, ANSTO -Access, services and case studies

The Australian Centre for Neutron Scattering at ANSTO is a multidisciplinary international centre of excellence, specialising in applying X-ray and neutron scattering techniques to the study of matter in various physical states; solid, liquid and gas. Over time, we've built an exceptional body of skills, experience and technical expertise, which we now offer to support industrial research and development.

The Australian Centre for Neutron Scattering Industrial Liaison Office (ACNS-ILO) was established in April 2014 to manage technology transfer and promote the use of our facilities in applied industrial research. Neutron characterisation can provides direct impact into the optimization of modern manufacturing processes, improved product reliability, enhanced design performance, reduced production cost, and extended life of significant engineering assets.

This presentation will showcase our flexibility in access and services. as well as highlight our recent achievements in helping industry across many sectors.

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